



A new class of antisense oligonucleotides targeting regions downstream of genes directs mRNA for degradation and decreases gene expression

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Background: Antisense oligonucleotides (ASOs) are short, chemically modified nucleic acid molecules designed to hybridize to target RNAs and thus modulate gene expression. Widely used ASO gapmers contain a central DNA region, which forms a heteroduplex with the target RNA, triggering RNase H1-dependent RNA degradation. **Method:** Using human cells, transcriptomics and molecular biology approaches, we investigated whether gapmer ASOs can act on nascent RNA synthesised in transcription termination windows located downstream of annotated genes. **Results:** We describe a new class of gapmer ASOs that expand the range of target sequences and reveal a critical blind spot in current ASO off-target assessment. We identify non-coding RNAs transcribed from transcription termination windows as previously unrecognized ASO targets. We show that downstream-of-gene ASOs (DG-ASOs) induce RNase H1-dependent cleavage, which impairs mRNA 3' end processing and ultimately directs the unprocessed mRNAs for degradation, leading to a pronounced decrease in expression of the corresponding upstream gene. **Conclusion:** This novel mode of antisense regulation uncovers downstream-of-gene termination windows as genuine ASO targets that can be exploited to suppress gene expression. Importantly, it also reveals an underappreciated source of off-target effects that may arise from ASOs binding in these regions. Our findings indicate the need to expand ASO design and offtarget assessment guidelines to include termination-window sequences to improve therapeutic efficacy and safety.